

Effects of *Lactobacillus rhamnosus* Strain GG in Pediatric Obesity-related Liver Disease

*Pietro Vajro, §Claudia Mandato, §Maria Rosaria Licenziati, *Adriana Franzese,
 ||Dino Franco Vitale, *Selvaggia Lenta, *Maria Caropreso,
 ‡Gianfranco Vallone, and †Rosaria Meli

ABSTRACT

Objective: Various lines of evidence suggest that malfunctioning of the gut–liver axis contributes to hepatic damage of rodents and humans with nonalcoholic fatty liver disease. We evaluated the effects of short-term probiotic treatment in children with obesity-related liver disease who were noncompliant with lifestyle interventions.

Patients and Methods: Twenty obese children (age 10.7 ± 2.1 years) with persisting hypertransaminasemia and ultrasonographic (US) bright liver were enrolled in this double-blind, placebo-controlled pilot study. At baseline, patients underwent clinical and laboratory anthropometric evaluation, measurement of the US hepatorenal ratio, standard liver function tests, oral glucose tolerance test, serum tumor necrosis factor- α , the glucose hydrogen breath test, and evaluation of serum antibodies to anti-peptidoglycan-polysaccharide polymers. After exclusion of causes of liver disease other than obesity, patients received either probiotic *Lactobacillus rhamnosus* strain GG (12 billion CFU/day) or placebo for 8 weeks.

Results: Multivariate analysis after probiotic treatment revealed a significant decrease in alanine aminotransferase (average variation vs placebo $P=0.03$) and in anti-peptidoglycan-polysaccharide antibodies (average variation vs placebo $P=0.03$) irrespective of changes in BMI z score and visceral fat. Tumor necrosis factor- α , and US bright liver parameters remained fairly stable.

Conclusions: Probiotic *L rhamnosus* strain GG warrants consideration as a therapeutic tool to treat hypertransaminasemia in hepatopathic obese children noncompliant with lifestyle interventions.

Key Words: children, *Lactobacillus* GG, liver, nonalcoholic fatty liver disease, probiotics

(JPGN 2011;52: 740–743)

Nonalcoholic fatty liver disease (NAFLD) is a worldwide medical problem that parallels the obesity epidemic (1) and has become the most common liver disorder at all ages (2,3). Loss of

weight and regular physical activity are the mainstay of treatment, but these are difficult to achieve (4–7). Although histological liver abnormalities are not generally a major concern in children, treatment is essential to avoid progression to more severe forms during or after transition into adult age (7,8). However, no proven effective pharmacological therapies are yet available (9,10).

Several studies have suggested that small intestinal bacterial overgrowth (SIBO) may contribute to the pathogenesis of NAFLD (11–13). Manipulation of intestinal flora with probiotics in experimental NAFLD of rodent animal models did indeed have a beneficial effect (14,15). In humans, a preliminary study investigating the outcome of probiotic VSL#3 (a mixture of 4 lactobacilli strains) therapy on liver dysfunction parameters in different categories of adult chronic liver disease including NAFLD also yielded promising results (16); however, the opposite results were reported in a recent article (17). The aim of our pilot study was, therefore, to evaluate whether a short course of therapy with probiotics improves the outcome of liver function tests in children with obesity-related liver disease.

PATIENTS AND METHODS

Obese children followed up at our pediatric obesity and pediatric liver units, with a body mass index (BMI) > 95th percentile for age and sex, who failed to adhere to previous slimming diets and who had not undergone previous pharmacological treatment for obesity, were offered the possibility of entering the present study. Twenty patients with persistent (>3 months) liver abnormalities (alanine aminotransferases [ALT] higher than upper normal values [40 U/L] associated with ultrasonographic [US] liver brightness suggestive of fatty liver) were enrolled.

Exclusion criteria were the coexistence of causes of increased transaminase levels other than obesity (namely muscular disease, viral hepatitis B and C, autoimmune hepatitis, α_1 -antitrypsin deficiency, cystic fibrosis, Wilson disease, hemochromatosis, hereditary fructose intolerance, amino acid disorders, atypical celiac disease, alcohol abuse, and drug toxicity), which were investigated by appropriate biochemical tests or verified by anamnestic data. Patients receiving concomitant antibiotic treatment were also excluded.

At entry, weight, height, BMI, BMI z score, and visceral fat thickness measured by US (18) were investigated. Standard liver function tests and US brightness were evaluated and graded as described elsewhere (4,19). A quantitative ratio of liver versus renal brightness that referred to specific regions of interest was obtained according to Mancini et al (20). All of the patients underwent a standard oral glucose tolerance test. The glucose H_2 breath test (H2BT) was performed after an overnight fast (1 g/kg, maximum 50 g/250 mL) (21). Patients were asked to avoid foods containing beans, bran, and high-fiber cereals on the day before the test. Breath

Received February 21, 2011; accepted April 13, 2011.

From the Departments of *Pediatrics, †Experimental Pharmacology, and ‡Radiology, University of Naples Federico II, the §Pediatria AORN Santobono, Naples, and the ||Fondazione Salvatore Maugeri IRCCS Istituto di Telese, Benevento, Italy.

Address correspondence and reprint requests to Prof Pietro Vajro, Dipartimento di Pediatria, Università di Napoli Federico II, Via Pansini 5, 80131 Napoli, Italy (e-mail: vajro@unina.it, pvajro@unisa.it).

This work was partly funded by the Italian Ministry of University and Research (MIUR) PRIN 2005.

The authors report no conflicts of interest.

Copyright © 2011 by European Society for Pediatric Gastroenterology, Hepatology, and Nutrition and North American Society for Pediatric Gastroenterology, Hepatology, and Nutrition

DOI: 10.1097/MPG.0b013e31821f9b85

samples were collected every 15 minutes for 2 hours. The test was considered positive for SIBO if the fasting breath H₂ concentration was > 20 ppm and/or increased > 10 ppm above baseline within the first 2 hours. Serum levels of tumor necrosis factor- α (TNF- α) were measured by quantitative sandwich enzyme immunoassay technique (R&D Systems, Minneapolis, MN). Serum antibodies to peptidoglycan-polysaccharide (PG-PS) complexes were determined by classical enzyme-linked immunosorbent assay according to the method of Hazenberg et al (22) and using commercially available PG-PS 100 P (Fluka Chemie GmbH, Buchs, Switzerland) (23). In the case of the hepatorenal US ratio, TNF- α , and PG-PS IgA, we compared the values in 20 age-, sex-, and weight-matched obese nonhepatopathic children.

The 20 children were randomly allocated to 1 of the 2 groups for a pilot randomized, double-blind (sealed envelopes), placebo-controlled study. The study drugs were oral *Lactobacillus GG* (12 billion CFU/day) and an indistinguishable placebo administered for 8 weeks. The primary outcome measure was improvement and/or normalization of ALT levels (< 40 IU/L). Secondary endpoints were treatment-related changes in liver echogenicity, TNF- α values, and, possibly, intestinal tests (H2BT and PG-PS antibodies).

Changes in lifestyle were not encouraged during the brief framework of the study. Parents were instructed to report any adverse effect that occurred during treatment and to supervise the administration of pills. Parents gave written informed consent. The study was conducted in accordance with Good Clinical Practices and the Declaration of Helsinki, and was approved by the ethics committee of our university hospital (number 8404).

Statistical Analysis

Continuous variables are reported as mean \pm standard deviation. Two-tailed tests of significance are reported, and $P < 0.05$ were considered statistically significant. The unpaired Student t test was used to assess differences in the average variations (8 weeks–baseline) between the treatment and placebo groups, as well as between the baseline means of the 2 groups. The possible confounding effect of BMI z score variations on ALT concentrations was explored by a multiple regression model with ALT as dependent variable and with BMI z score variation and treatment as independent variables. The Fisher exact test was used to calculate an exact probability value for the relation between 2 dichotomous variables.

RESULTS

Demographic data, anthropometric measurements, and biochemical and ultrasonographic features in the 20 patients at entry into the study are shown in Table 1. Baseline results confirmed that isolated hypertransaminasemia was the only abnormality to emerge from the liver tests. All of the children had mild to moderate US fatty liver. Diabetes was excluded in all of the children based on the results of oral glucose tolerance test. H2BT was abnormal (H₂ peak of 19 ppm above baseline at 60 minutes) only in 1 patient (of the treated group).

All of the patients completed the 8-week study with good adherence to therapy as documented by pill count and parents' supervision reports. No adverse effects were reported. Anthropometric, laboratory, and instrumental findings in the 2 groups of patients at baseline and after 8 weeks of probiotic or placebo treatment are listed in Table 2. As depicted also in Figure 1, the average variations of ALT levels in the 2 arms of the study were significantly more pronounced in the *Lactobacillus GG*–treated group ($P = 0.03$). Eight of the 10 treated patients and 3 of 10 patients in the placebo group attained ALT values < 40 U/L, but the difference did not reach statistical significance ($P = 0.069$ Fisher

TABLE 1. Demographic data, anthropometric measurements, and biochemical and ultrasonographic features in the 20 hepatopathic obese children at study entry

	Patients, n = 20
Sex, M/F	18/2
Age, y	10.7 \pm 2.1
Weight, kg	61.7 \pm 12.7
Height, cm	148.0 \pm 10.0
BMI z score	2.2 \pm 0.27
US visceral fat, mm	11.4 \pm 2.9
ALT, IU/L (normal values < 40)	66.9 \pm 27.3
AST, IU/L (normal values < 40)	41.6 \pm 16.54
Triglycerides, mg/dL	96.85 \pm 9.7
Cholesterol, mg/dL	157.0 \pm 29.9
PG-PS IgA, OD*	0.50 \pm 0.20 [†]
TNF- α , pg/mL*	8.04 \pm 5.0 [‡]
Hepatorenal US ratio*	1.24 \pm 0.21 [§]

ALT = alanine aminotransferase; AST = aspartate aminotransferase; BMI = body mass index; IgA = immunoglobulin A; OD = optical density; PG-PS = peptidoglycan-polysaccharide; TNF- α = tumor necrosis factor- α ; US = ultrasonographic.

*Normal values in 20 age-, sex-, and weight-matched nonhepatopathic obese children: [†]0.33 \pm 0.11 ($P < 0.008$); [‡]7.27 \pm 3.11 ($P = NS$); [§]1.10 \pm 0.11 ($P = 0.003$).

exact test). PG-PS IgA significantly decreased only in the probiotic-treated group. Anthropometric data including US visceral fat, TNF- α , and US liver brightness evaluated by the hepatorenal ratio remained fairly stable in both groups during the study (Table 2).

DISCUSSION

This is the first study to investigate the effects of probiotic treatment on obesity-related liver abnormalities in children. The study was designed as a short-term pilot trial to circumvent the frequently reported unpredictable effects of lifestyle changes that usually confound the results of a long-term study in this population. In fact, even minimal weight and lifestyle changes may affect the biochemical parameters of NAFLD (4–6). Here we show that a short course of probiotic treatment significantly improved ALT values, which in most cases became normal, and that this effect was independent of weight changes.

Small intestinal bacterial overgrowth has been implicated in the pathogenesis of NAFLD (11–13). The results of the glucose H2BT, which is a surrogate test for SIBO evaluation, in our children did not confirm the increased prevalence of SIBO reported in morbidly obese adults (12). However, the level of PG-PS IgA antibodies, another surrogate for SIBO, was significantly higher than that of a group of 20 age-, sex-, and weight-matched obese nonhepatopathic controls (Table 1), and moreover they significantly decreased only in the children treated with probiotics and not in the placebo-treated group. Because PG-PS polymers are integral cell wall components of most bacterial species, including intestinal Bacteroides, which are the major determinants of SIBO (24,25), the decrease of PG-PS IgA antibodies after treatment may imply abnormalities in the patients' small intestinal microflora. Wigg et al (11) found that levels of endotoxin (ie, another SIBO marker) did not differ between patients with NAFLD and healthy controls. In a more recent work, endotoxin levels in the peripheral circulation were increased in patients with NAFLD, but they were unrelated to disease severity (26). Given the differences between

TABLE 2. Anthropometric, laboratory, biochemical, and instrumental findings in the 2 groups of children with obesity-related liver disease at baseline (T0) and after 8 weeks of probiotic or placebo treatment (T8)

	Controls (placebo)		Probiotic treatment		Average variation (probiotic vs placebo)	Baseline <i>t</i> test
	T0 baseline	T8 weeks	T0 baseline	T8 weeks		
ALT, IU/L	63.6 ± 18.47	61.6 ± 31.80	70.3 ± 34.76	40.1 ± 22.37	0.03	NS
BMI <i>z</i> score	2.12 ± 0.24	2.00 ± 0.26	2.29 ± 0.30	2.21 ± 0.31	NS	NS
Visceral fat, mm	10.79 ± 3.29	11.39 ± 2.35	12.04 ± 2.58	14.04 ± 5.14	NS	NS
TNF- α , pg/mL	8.28 ± 5.11	6.88 ± 6.43	7.74 ± 5.28	5.42 ± 2.10	NS	NS
Hepatorenal US ratio	1.31 ± 0.26	1.30 ± 0.15	1.17 ± 0.12	1.22 ± 0.12	NS	NS
PG-PS IgA, OD	0.64 ± 0.32	0.68 ± 0.36	0.68 ± 0.26	0.60 ± 0.25	0.03	NS

ALT = alanine aminotransferase; BMI = body mass index; IgA = immunoglobulin A; NS = nonsignificant; OD = optical density; PG-PS = peptidoglycan-polysaccharide; TNF- α = tumor necrosis factor-alpha; US = ultrasonographic.

peripheral and portal blood values and potential methodological difficulties caused by the short half-life of endotoxins, data obtained from the measurement of serum endotoxin levels should be interpreted with caution (11,26). For this reason, we did not measure endotoxin levels in the present study.

TNF- α is believed to be one of the protagonists of the inflammatory response in NAFLD after stimulation via Toll-like receptors by endotoxins, which reach the liver via portal flow through a more permeable intestinal barrier. However, similar to a previous study (11), the TNF- α values in our hepatopathic obese patients overlapped those of the 20 age- and weight-paired obese individuals. They tended toward a mild decrease in both treatment arms without, however, reaching statistical significance. *Lactobacillus* GG-induced modulation on studied parameters involved in NAFLD pathogenesis will probably require long-term studies. The effects of probiotics on NAFLD have been evaluated in animal models and, although to a lesser extent, in human adults. Probiotic VSL#3 treatment in NAFLD ob/ob mice lowered the transaminase level and affected histological liver inflammation without modifying steatosis (14). The same treatment in mice with diet-induced NAFLD attenuated fibrosis without affecting steatohepatitis (15,27). The unchanged US hepatorenal ratios in our patients are in line with these data. Another study showed that probiotics had a beneficial effect on several types of human liver disease including

NAFLD, probably acting on the steatosis component (16). Contrary to what may be expected from the results of animal studies, a recent preliminary report on human NAFLD found a reversible increase in magnetic resonance imaging-calculated fat liver content during probiotic treatment (17). However, details of possible simultaneous weight changes were not reported. Given the above, individuals enrolled in such studies should undergo accurate anthropometric monitoring to ensure that other factors do not confound the effect of treatment on the outcome.

Our results do not shed light on the mechanism(s) underlying the improvement of our patients' ALT values. Recent evidence indicates that there are quantitative (ie, SIBO) and qualitative differences in gut microbiota and microbiome between lean and obese individuals (28). The fact that our patients were completely asymptomatic from a gastrointestinal viewpoint may argue against the existence of a true SIBO. Because positive H2BTs in the exceedingly large percentage of Wiggs' patient series has been questioned by several investigators (29), it is feasible that in our study, *Lactobacillus* GG exerted a beneficial effect on obesity-related liver disease by modulating an altered bacterial composition rather than reducing its quantity (30). However, methodological aspects may have affected the results of the glucose H2BT in detecting SIBO; namely, the dose of the sugar or the duration of the test may not have been sufficient to exclude a bacterial contamination restricted to the distal small bowel. (21) Future large-scale intervention trials investigating the potential benefit of probiotics should include a combination of gastrointestinal dysfunction tests, including lactulose H2BT and intestinal permeability studies.

In view of the excellent tolerance and the remarkable reduction of transaminase after probiotics in our double-blind, placebo-controlled, short-term pilot study, *Lactobacillus rhamnosus* strain GG could be considered a potential therapeutic tool for pediatric obesity-related liver disease in children who are unable to lose weight. Our promising data provide a rationale for starting a larger study on possibly biopsy-proven pediatric NAFLD.

Acknowledgments: We are grateful to Patrizia Colicchio, MD, and Brunella Aceto, dietician, for assistance in the clinical management of patients; Maria Passaretti, PhD, and Luciano Rapacciuolo, PhD, for laboratory assistance; and Francesco Manguso, MD, for critical evaluation of the manuscript. We are indebted to Jean Ann Gilder (Scientific Communication srl) for editing the text. *Lactobacillus rhamnosus* strain GG (ATCC 53103)

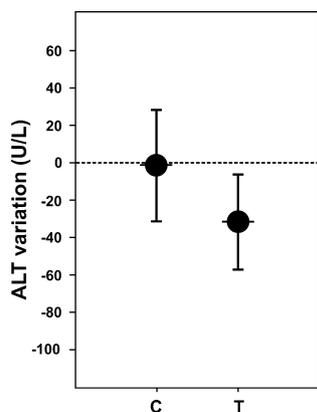


FIGURE 1. Effects of treatment on serum alanine aminotransferase (ALT) values in patients receiving either placebo (placebo controls, C) or *Lactobacillus* GG (T). $P = 0.03$.

with verified composition and indistinguishable placebo were supplied by Dicofarm SpA, Rome, Italy.

REFERENCES

1. Yanovski SZ, Yanovski JA. Obesity prevalence in the United States—up, down, or sideways? *N Engl J Med* 2011;364:987–9.
2. Younossi ZM, Stepanova M, Afendy M, et al. Changes in the prevalence of the most common causes of chronic liver diseases in the United States from 1988 to 2008. *Clin Gastroenterol Hepatol*. 2011 Apr 11 [Epub ahead of print].
3. Schwimmer JB, Deutsch R, Kahen T, et al. Prevalence of fatty liver in children and adolescents. *Pediatrics* 2006;118:1388–93.
4. Franzese A, Vajro P, Argenziano A, et al. Liver involvement in obese children. Ultrasonography and liver enzyme levels at diagnosis and during follow-up in an Italian population. *Dig Dis Sci* 1997;42:1428–32.
5. St George A, Bauman A, Johnston A, et al. Effect of a lifestyle intervention in patients with abnormal liver enzymes and metabolic risk factors. *J Gastroenterol Hepatol* 2009;24:399–407.
6. Nobili V, Alisi A, Raponi M. Pediatric non-alcoholic fatty liver disease: preventive and therapeutic value of lifestyle intervention. *World J Gastroenterol* 2009;15:6017–22.
7. Bellentani S, Scaglioni F, Marino M, et al. Epidemiology of non-alcoholic fatty liver disease. *Dig Dis* 2010;28:155–61.
8. Caldwell S, Argo C. The natural history of non-alcoholic fatty liver disease. *Dig Dis* 2010;28:162–8.
9. Musso G, Gambino R, Cassader M, et al. A meta-analysis of randomized trials for the treatment of nonalcoholic fatty liver disease. *Hepatology* 2010;52:79–104.
10. Socha P, Horvath A, Vajro P, et al. Pharmacological interventions for nonalcoholic fatty liver disease in adults and in children: a systematic review. *J Pediatr Gastroenterol Nutr* 2009;48:587–96.
11. Wigg AJ, Roberts-Thomson IC, Dymock RB, et al. The role of small intestinal overgrowth, intestinal permeability, endotoxaemia, and tumor necrosis factor α in the pathogenesis of non alcoholic steatohepatitis. *Gut* 2001;48:206–11.
12. Sabaté JM, Jouët P, Harnois F, et al. High prevalence of small intestinal bacterial overgrowth in patients with morbid obesity: a contributor to severe hepatic steatosis. *Obes Surg* 2008;18:371–7.
13. Miele L, Valenza V, La Torre G, et al. Increased intestinal permeability and tight junction alterations in nonalcoholic fatty liver disease. *Hepatology* 2009;49:1877–87.
14. Li Z, Yang S, Lin H, et al. Probiotics and antibodies to TNF inhibit inflammatory activity and improve nonalcoholic fatty liver disease. *Hepatology* 2003;37:343–50.
15. Esposito E, Iacono A, Bianco G, et al. Probiotics reduce the inflammatory response induced by a high-fat diet in the liver of young rats. *J Nutr* 2009;139:905–11.
16. Loguercio C, Federico A, Tuccillo C, et al. Beneficial effects of a probiotic VSL#3 on parameters of liver dysfunction in chronic liver diseases. *J Clin Gastroenterol* 2005;39:540–3.
17. Solga SF, Buckley G, Clark JM, et al. The effect of a probiotic on hepatic steatosis. *J Clin Gastroenterol* 2008;42:1117–9.
18. Sabir N, Sermez Y, Kazil S, et al. Correlation of abdominal fat accumulation and liver steatosis: importance of ultrasonographic and anthropometric measurements. *Eur J Ultrasound* 2001;14:121–8.
19. Vajro P, Fontanella A, Perna C, et al. Persistent hyperaminotransferasemia resolving after weight reduction in obese children. *J Pediatr* 1994;125:239–41.
20. Mancini M, Prinster A, Annuzzi G, et al. Sonographic hepatic-renal ratio as indicator of hepatic steatosis: comparison with (1)H magnetic resonance spectroscopy. *Metabolism* 2009;58:1724–30.
21. Gasbarrini A, Corazza GR, Gasbarrini G, et al. Methodology and indications of H2-breath testing in gastrointestinal diseases: the Rome Consensus Conference. *Aliment Pharmacol Ther* 2009;29(suppl 1):1–49.
22. Hazenberg MP, de Visser H, Bras MJ, et al. Serum antibodies to peptidoglycan-polysaccharide complexes from the anaerobic intestinal flora in patients with Crohn's disease. *Digestion* 1990;47:172–80.
23. Schrijver IA, De Man YA, Melief MJ, et al. Reduced systemic IgG levels against peptidoglycan in rheumatoid arthritis (RA) patients. *Clin Exp Immunol* 2001;123:140–6.
24. Lichtman SN, Keku J, Schwab JH, et al. Evidence for peptidoglycan absorption in rats with experimental small bowel bacterial overgrowth. *Infect Immun* 1991;59:555–62.
25. Lumsden AB, Henderson JM, Kutner MH. Endotoxin levels measured by a chromogenic assay in portal, hepatic and peripheral venous blood in patients with cirrhosis. *Hepatology* 1988;8:232–6.
26. Harte AL, da Silva NF, Creely SJ, et al. Elevated endotoxin levels in non-alcoholic fatty liver disease. *J Inflamm (Lond)* 2010;7:15.
27. Velayudham A, Dolganiuc A, Ellis M, et al. VSL#3 probiotic treatment attenuates fibrosis without changes in steatohepatitis in a diet-induced nonalcoholic steatohepatitis model in mice. *Hepatology* 2009;49:989–97.
28. Turnbaugh PJ, Gordon JI. The core gut microbiome, energy balance and obesity. *J Physiol* 2009;587:4153–8.
29. Riordan SM, Duncombe VM, Thomas MC, et al. Small intestinal bacterial overgrowth, intestinal permeability, and non-alcoholic steatohepatitis. *Gut* 2002;50:136–8.
30. Abu-Shanab A, Quigley EM. The role of the gut microbiota in non-alcoholic fatty liver disease. *Nat Rev Gastroenterol Hepatol* 2010;7:691–701.